

FORM PTO-1390	U.S. Department of Commerce Patent and Trademark Office	Attorney's Docket No.
TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371		1871-133
		U.S. Application No. (if known, see 37 CFR 1.5) 10/049759
INTERNATIONAL APPLICATION NO. PCT/AU00/00980	INTERNATIONAL FILING DATE August 16, 2000	PRIORITY DATE CLAIMED August 16, 1999
TITLE OF INVENTION Methods for Diagnosis and Treatment of Human Disease Including Hypertension		
APPLICANT(S) FOR DO/EO/US David Ian COOK, Kristie-Ann FRALEY, Hajime ISHIBASHI, Permsak KOMWATANA, Angeles SANCHEZ-PEREZ, John YOUNG, Anuwar DINUDOM		
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:		
<ol style="list-style-type: none"> 1. <input checked="" type="checkbox"/> This is a FIRST submission of items concerning a filing under 35 U.S.C. 371 2. <input type="checkbox"/> This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. 3. <input type="checkbox"/> This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (21) indicated below. 4. <input type="checkbox"/> The US has been elected by the expiration of 19 months from the priority date (Article 31). 5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371(c)(2)) <ol style="list-style-type: none"> a. <input type="checkbox"/> is attached hereto (required only if not communicated by the International Bureau). b. <input checked="" type="checkbox"/> has been communicated by the International Bureau. c. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US) 6. <input type="checkbox"/> An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)). <ol style="list-style-type: none"> a. <input type="checkbox"/> is attached hereto. b. <input type="checkbox"/> has been previously submitted under 35 U.S.C. 154(d)(4). 7. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)) <ol style="list-style-type: none"> a. <input type="checkbox"/> are attached hereto (required only if not communicated by the International Bureau). b. <input type="checkbox"/> have been communicated by the International Bureau. c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired. d. <input checked="" type="checkbox"/> have not been made and will not be made. 8. <input type="checkbox"/> An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)). 9. <input type="checkbox"/> An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). 10. <input type="checkbox"/> An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)). 		
ITEMS 11. TO 20. below concern other document(s) or information included:		
<ol style="list-style-type: none"> 11. <input type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98. 12. <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included. 13. <input checked="" type="checkbox"/> A FIRST preliminary amendment. 14. <input type="checkbox"/> A SECOND or SUBSEQUENT preliminary amendment. 15. <input type="checkbox"/> A substitute specification. 16. <input type="checkbox"/> A change of power of attorney and/or address letter. 17. <input type="checkbox"/> A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821-1.825 18. <input type="checkbox"/> A second copy of the published international application under 35 U.S.C. 154(d)(4). 19. <input type="checkbox"/> A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4). 20. <input checked="" type="checkbox"/> Other items or information: <ul style="list-style-type: none"> - Published application WO 01/12805 with Search Report - International Preliminary Examination Report with amended pages submitted under Article 34 - Application Data Sheet 		

U.S. APPLICATION NO. (If known, see 37 CFR 1.50) 10/049759		INTERNATIONAL APPLICATION NO. PCT/AU00/00980		ATTORNEY DOCKET NO. 1871-133	
21. <input checked="" type="checkbox"/> The following fees are submitted: Basic National Fee (37 CFR 1.492)(a)(1)-(5): Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report Not Prepared by EPO or JPO. \$ 1,040.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report has been prepared by the EPO or JPO. \$ 890.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO. \$ 740.00 International preliminary examination fee (37 CFR 1.482) paid to USPTO but claims did not satisfy provisions of PCT Article 33(1)-(4). \$ 710.00 International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4). \$ 100.00				<u>CALCULATIONS</u>	<u>PTO USE ONLY</u>
ENTER APPROPRIATE BASIC FEE AMOUNT =				\$ 1,040.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than [] 20 [] 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$	
Claims	Number Filed	Number Extra	Rate		
Total Claims	59 -20 =	39	X \$18.00	\$ 702.00	
Independent Claims	13 - 3 =	10	X \$84.00	\$ 840.00	
Multiple dependent claim(s) (if applicable)				+ \$280.00	\$ 280.00
TOTAL OF ABOVE CALCULATIONS =				\$ 2,862.00	
<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				\$	
SUBTOTAL =				\$ 2,862.00	
Processing fee of \$130.00 for furnishing the English translation later than [] 20 [] 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				+	\$
TOTAL NATIONAL FEE =				\$ 2,862.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property				+	\$
TOTAL FEES ENCLOSED =				\$ 2,862.00	
				Amount to be refunded	\$
				charged	\$
a. <input type="checkbox"/>	A check in the amount of \$ _____ to cover the above fees is enclosed.				
b. <input checked="" type="checkbox"/>	Please charge my Deposit Account No. 02-2135 in the amount of \$ <u>2,862.00</u> to cover the above fees. A duplicate copy of this sheet is enclosed.				
c. <input checked="" type="checkbox"/>	The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 02-2135. A duplicate copy of this sheet is enclosed.				
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO:			 Signature Barbara G. Ernst Name 30,377 Registration Number		
Customer No. 6449					
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<p>IN THE UNITED STATES PATENT AND TRADEMARK OFFICE</p>	<i>Application Number</i>	PCT/AU00/00980
	<i>Filing Date</i>	August 16, 2000
	<i>First Named Inventor</i>	David Ian COOK
	<i>Group Art Unit</i>	Unassigned
	<i>Examiner Name</i>	Unassigned
	<i>Attorney Docket Number</i>	1871-133
<i>Title of the Invention:</i> METHODS FOR DIAGNOSIS AND TREATMENT OF HUMAN DISEASE INCLUDING HYPERTENSION		

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
Washington, D.C. 20231

Dear Sir:

Preliminary to examination on the merits, please amend the above-referenced application as follows:

Amend claims 16, 20, 21, 32, and 37 as shown on the following pages.

Marked-up copies of the original text of the amended claims, specification portions and abstract are attached to this amendment. Material inserted is indicated by redlining (insertion) and material deleted is indicated by strike-out (~~strike-out~~).
therefor.

Clean Copy of Amended Claims

16. (Amended) A method according to claim 13 or 14, wherein the sample of cells is assessed for reduced or over activity of the Na^+ transport protein by determining the rate of Na^+ -dependent intracellular pH (pH) recovery and comparing said rate against similarly measured rates from cells from healthy tissue isolated from said subject or a control subject(s).

20. (Amended) A method according to claim 17, 18 or 19, wherein said Na^+ transport protein is selected from the group consisting of Na^+ - H^+ exchanger 1 (NHE1), Na^+ - H^+ exchanger 2 (NHE2), Na^+ - H^+ exchanger 3 (NHE3), the Na^+ - HCO_3 cotransporter and the Na^+ K^+ 2Cl^- cotransporter.

21. (Amended) A method according to claim 1, 8, 13, 17, 18 or 19, wherein the said human disease is selected from hypertension, renal failure, cardiac hypertrophy and cardiological syndrome X.

32. (Amended) A host cell transformed with a DNA molecule according to claim 22 or 37.

37. (Amended) A method for detecting agonist or antagonist agents of the receptor of claim 34 or 35, wherein said method comprises contacting said receptor, or a host cell transformed with and expressing the DNA molecule of claim 22 or 27, with a test agent under conditions enabling the activation of said receptor, and detecting an increase or decrease in activity of the receptor.

REMARKS

The amendments to the claims are made to correct improper claim dependencies and to put the claims in the format preferred by the U.S. Patent Office. No new matter is introduced by means of these amendments.

RESPECTFULLY SUBMITTED,					
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Attachments: Marked-Up Copies of Amendments to the Claims

Amended Claims: Version with markings to show changes made

16. (Amended) A method according to ~~any one of claims 13 to 15~~ ~~claim 13 or 14~~, wherein the sample of cells is assessed for reduced or over activity of the Na^+ transport protein by determining the rate of Na^+ -dependent intracellular pH (pH) recovery and comparing said rate against similarly measured rates from cells from healthy tissue isolated from said subject or a control subject(s).

20. (Amended) A method according to ~~any one of claims 17-19~~ ~~claim 17, 18 or 19~~, wherein said Na^+ transport protein is selected from the group consisting of Na^+ - H^+ exchanger 1 (NHE1), Na^+ - H^+ exchanger 2 (NHE2), Na^+ - H^+ exchanger 3 (NHE3), the Na^+ - HCO_3 cotransporter and the Na^+ K^+ 2Cl cotransporter.

21. (Amended) A method according to ~~any one of claims 1 to 20~~ ~~claim 1, 8, 13, 17, 18 or 19~~, wherein the said human disease is selected from hypertension, renal failure, cardiac hypertrophy and cardiological syndrome X.

32. (Amended) A host cell transformed with a DNA molecule according to ~~any one of claims 22 to 31~~ ~~claim 22 or 37~~.

37. (Amended) A method for detecting agonist or antagonist agents of the receptor of claim 34 or 35, wherein said method comprises contacting said receptor, or a host cell transformed with and expressing the DNA molecule of ~~any one of claims 22 to 31~~ ~~claim 22 or 27~~, with a test agent under conditions enabling the activation of said receptor, and detecting an increase or decrease in activity of the receptor.

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METHODS FOR DIAGNOSIS AND TREATMENT OF HUMAN DISEASES
INCLUDING HYPERTENSION

Field of the Invention:

5 The present invention relates to the diagnosis and treatment of human disease, particularly human disease characterised by abnormal cytosolic ion composition resulting from reduced or over activity of Na^+ transport proteins such as the ubiquitous Na^+ - H^+ exchanger, NHE1.

10 Background of the Invention:

In recent years, over activity of Na^+ transporting systems in absorptive epithelia has been implicated in the pathogenesis of a number of major diseases including hypertension (1, 2), diabetic nephropathy (3), cardiological syndrome X (4), ventricular hypertrophy (5), chronic pulmonary hypertension (6) and cystic fibrosis (7). In the case of the hereditary hypertensive disease known as Liddle's syndrome, this activity has been attributed to a mutation of the epithelial Na^+ channel (ENaC) leading to loss of an inhibitory feedback mechanism which normally switches off Na^+ channel activity in response to increased intracellular Na^+ (8, 9). The mechanisms that underlie this so-called homocellular regulation have been the subject of controversy, but recent experiments have revealed a previously unsuspected mechanism in which cytosolic Na^+ is "sensed" by an intracellular receptor (10). This receptor activates the G protein, G_o (11), the α -subunit of which then causes the ubiquitin-protein ligase, Nedd4 (10), to ubiquitinate and inactivate the epithelial Na^+ channels (12, 13). This receptor for intracellular Na^+ is blocked by amiloride and amiloride analogs such as dimethylamiloride and benzimidazole guanidinium (10), thus explaining the previously puzzling ability of these agents to stimulate Na^+ channel activity (14).

20 The present applicants have now found that the intracellular Na^+ receptor that controls absorptive epithelial Na^+ channels also controls the activity of the ubiquitous isoform of the Na^+ - H^+ exchanger 1, NHE1 (34). This finding suggests that intracellular Na^+ receptors form part of a general mechanism for regulating Na^+ transport proteins. It is therefore anticipated that the intracellular Na^+ receptors (and the signal-transduction systems by which they control Na^+ channels, Na^+ - H^+ exchangers and other Na^+

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transporting proteins) shall provide a useful target for diagnostic assays and treatments for hypertension and other diseases.

Disclosure of the Invention:

5 Thus, in a first aspect, the present invention provides a method of treatment of a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport

10 protein has reduced activity, the method comprising administering to a subject having said disease an effective amount of an agent that blocks said inhibitory feedback mechanism and thereby substantially restores the ion composition of the cytosol in said diseased cells to that which is found in corresponding cells from healthy tissue.

15 Preferably, the present invention provides a method of treatment of a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from reduced activity of a Na^+ transport protein selected from those which are inactivated by ubiquitination (e.g. through the action of a ubiquitin-protein ligase) and, particularly, from those included in 20 the group consisting of NHE1, Na^+ - H^+ exchanger 2 (NHE2) (35), Na^+ - H^+ exchanger 3 (NHE3) (36), the Na^+ - HCO_3^- cotransporter (37) and the Na^+ K^+ 2Cl^- cotransporter (38).

25 Where the characteristic abnormal cytosolic ion composition arises from reduced Na^+ transport protein activity resulting from inappropriate activity of the Na^+ transport protein inhibitory feedback mechanism, the administered agent may be selected from agents capable of blocking the Na^+ transport protein inhibitory feedback mechanism. Preferred agents of the latter kind are amiloride and amiloride analogs (e.g. 6-iodoamiloride, N-dimethylamiloride, and benzimidazoylguanidium), G-protein inhibitors (e.g. 30 GDP- β -S (39) and NF023 (40)) and agents that inhibit the action of ubiquitin protein ligase on the Na^+ transport protein. Examples of this latter kind of agents are dominant negative mutants of ubiquitin (e.g. K48R (24)), agents that prevent binding of the ubiquitin protein ligase to the Na^+ transport protein (e.g. membrane permeable peptide analogs of the protein motif to 35 which the ubiquitin protein ligase binds such as the WW2 and WW3 domains of Nedd4 (10)), agents that prevent ubiquitination of the Na^+ transport protein

[e.g. membrane permeable peptide analogs of the protein motif which is actually ubiquitinated, such as the N-terminal of the α - or γ -subunit of ENaC (41)] and inhibitors of the effectors of ubiquitin action on the Na^+ transport protein including proteins involved in endocytosis (e.g. membrane permeable analogs of amphiphysin SH3 peptide(42)), and inhibitors of the degradation of the Na^+ transport protein by proteasomes (e.g. lactacystin) or lysosomes (e.g. bafilomycin or chloroquine). Peptide analogs may be made to be membrane permeant by including a *Drosophila* antennapedia homeobox domain (15, 18).

10 In a second aspect, the present invention provides a method of treatment of a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from the loss or inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has over activity, the method comprising administering to a subject having said disease an effective amount of an agent that substantially restores the ion composition of the cytosol in said diseased cells to that which is found in corresponding cells from healthy tissue.

Where the characteristic abnormal cytosolic ion composition arises from Na^+ transport protein over activity resulting from, for example, loss of the Na^+ transport protein inhibitory feedback mechanism or inappropriate activity of other control systems (e.g. excessive levels of growth factors or glucose), the administered agent may be selected from gene therapy agents (e.g. adenoviruses capable of causing the expression of a protein participating in the Na^+ transport protein inhibitory feedback mechanism), intracellular Na^+ receptor activators (e.g. guanidium and guanidium analogs), G-protein activators (e.g. GTP- γ -S (43) and receptor mimetic peptides such as APP20(17)), ubiquitin ligase activators (e.g. membrane permeable peptides that mimic the effect of active G proteins on the ubiquitin protein ligase), and agents that trigger endocytosis.

An "effective amount" of the agent used in the method of the first or second aspect will depend upon the particular agent used, however, generally, the amount would be expected to be below about 10 mg/kg. For example, an effective amount of amiloride or an amiloride analog would typically be in the range of 1 to 3 mg/kg.

The agent may be formulated with various pharmaceutically-acceptable excipients and/or carriers commonly used in the art and prepared for administration orally (e.g. as tablets, capsules, caplets or liquids), nasally (e.g. aerosol sprays), rectally (e.g. as suppositories) and transdermally (e.g. as a transdermal patch or dermally absorbed cream or lotion). The agent may also be formulated as an injectible solution or suspension for subcutaneous, intravenous or intramuscular administration.

In a third aspect, the present invention provides a method of diagnosis of a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from the loss of or inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has reduced or over activity, the method comprising isolating from a subject suspected of having said disease a sample of cells (such as epithelial cells or lymphocytes) and assessing said sample of cells for reduced or over activity of said Na^+ transport protein resulting from the loss of an inappropriate activity of said inhibitory feedback mechanism or otherwise assessing said sample of cells for the loss of or inappropriate activity of said inhibitory feedback mechanism.

Preferably, the present invention provides a method of diagnosis of a human disease which is characterised by abnormal cytosolic ion composition resulting from reduced or over activity of a Na^+ transport protein selected from those which are inactivated by ubiquitination (e.g. through the action of a ubiquitin-protein ligase) and, particularly, from those included in the group consisting of NHE1, NHE2, NHE3, the $\text{Na}^+ \text{-HCO}_3^-$ cotransporter and the $\text{Na}^+ \text{K}^+ \text{2Cl}^-$ cotransporters.

The sample of cells may be assessed for reduced or over activity of Na^+ transport protein by, for example, determining the rate of Na^+ -dependent intracellular pH (pH_i) recovery and comparing the value against similarly measured values from cells from healthy tissue isolated from the said suffering subject or from a control (i.e. non-diseased) subject or subjects (e.g. an average value from a panel of two or more healthy subjects).

In a variation of the invention according to the third aspect, the sample of diseased cells may be assessed for over or under expression of a protein participating in the Na^+ transport protein inhibitory feedback mechanism

(e.g. by polymerase chain (PCR) techniques, Northern blot hybridisation, Western blot or immunoprecipitation).

In a fourth aspect, the present invention provides a method of diagnosis of a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from the loss or inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has reduced or over activity, the method comprising isolating a genomic DNA sample from a subject suspected of having said disease and assessing said sample for the presence of a gene encoding a mutated product causitive of said reduced or over activity of said Na^+ transport protein.

In a fifth aspect, the present invention provides a method of assessing a subject for a predisposition to a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from the loss or inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has reduced or over activity, the method comprising isolating a genomic DNA sample from a subject and assessing said sample for the presence of a gene encoding a mutated product causitive of reduced or over activity of said Na^+ transport protein.

In the methods of the fourth and fifth aspects, the human disease is preferably one which is characterised by abnormal cytosolic ion composition resulting from reduced or over activity of a Na^+ transport protein which has been selected from the group consisting of NHE1, NHE2, NHE3, the Na^+ - HCO_3^- cotransporter and the $\text{Na}^+\text{K}^+2\text{Cl}^-$ cotransport protein. The genomic DNA sample may be isolated using routine protocols known to the art. The genomic DNA sample may be isolated from any cell sample such as whole blood, tissue biopsy or cheek cell sample. The assessment of the presence of a gene encoding a mutated product causitive of reduced or over activity of the Na^+ transport protein, may be preferably achieved by hybridisation or PCR techniques using probes/primers designed to specifically hybridise to genes including mutated nucleotide sequences. The gene whose presence is to be assessed may encode a mutated protein participating in the Na^+ transport protein inhibitory feedback mechanism (e.g. a mutated G-protein or mutated intracellular Na^+ receptor).

The methods of the invention are applicable to, for example, hypertension, renal failure, cardiac hypertrophy and cardiological syndrome X.

The present applicants have also found that the intracellular Na^+ receptor controlling NHE1 is blocked by amiloride and amiloride analogs with the following order of potency:

5 6-iodoamiloride ($\text{EC}_{50} = 0.1 \mu\text{mol/l}$) < amiloride ($1.0 \mu\text{mol/l}$)
< 5-N-dimethylamiloride ($30 \mu\text{mol/l}$), benzamil ($> 30 \mu\text{mol/l}$) <
benzimidazolylguanidium ($300 \mu\text{mol/l}$)

10 Knowledge of these differing potencies enables the isolation of a DNA molecule encoding the intracellular Na^+ receptor controlling NHE1. That is, by using the α -subunit of G_α as "bait" in a yeast two-hybrid technique ("The yeast two-hybrid system" edited by P.L. Bartel & S. Fields, Oxford University Press, Oxford, 1997), DNA molecules encoding interacting proteins may be

15 isolated from suitable cDNA or genomic DNA libraries and then screened for the ability of the encoded proteins to bind 6-iodoamiloride. Further screens may be conducted for the relative inability of the encoded proteins to bind benzamil, the ability of antibodies raised to the encoded proteins to immunoprecipitate the α -subunit of G_α , and the ability of antibodies raised to

20 the encoded proteins to block the NHE1 inhibitory feedback mechanism.

By using the yeast two-hybrid system with a constitutively active mutant of the α -subunit of G_α , it is possible to identify and isolate proteins which interact with active G_α and hence are involved in the inhibitory feedback mechanism at a loci downstream of G_α . Similarly, by using the

25 yeast two-hybrid system with a dominant negative mutant of the α -subunit of G_α , it is possible to identify and isolate proteins such as the intracellular Na^+ receptors which are involved in the inhibitory feedback mechanism at a loci upstream of G_α .

The present applicants have isolated 5 cDNA molecules from mouse

30 kidney and mandibular gland cDNA libraries encoding likely intracellular Na^+ receptors controlling NHE1 and Na^+ channels. The 5 candidates are nucleobindin (18), GAIP (19), rap1GAP (20) and novel proteins designated GILT (formerly designated c10a) and SCunique.

Thus, in a sixth aspect, the present invention provides an isolated DNA molecule encoding an intracellular Na^+ receptor designated GILT, said DNA molecule comprising a nucleotide sequence substantially corresponding to

Thus, in a sixth aspect, the present invention provides an isolated DNA molecule encoding an intracellular Na^+ receptor designated GILT, said DNA molecule comprising a nucleotide sequence substantially corresponding to that shown as SEQ ID NO: 1 or a nucleotide sequence showing $\geq 75\%$ (more preferably $\geq 85\%$, most preferably $\geq 95\%$) homology to that shown as SEQ ID NO: 1.

Preferably, the isolated DNA molecule of the sixth aspect encodes a protein comprising an amino acid sequence substantially corresponding to that shown as SEQ ID NO: 2.

10 In a seventh aspect, the present invention provides an isolated DNA molecule encoding an intracellular Na^+ receptor designated SCunique, said DNA molecule comprising a nucleotide sequence substantially corresponding to that shown as SEQ ID NO: 3 or a nucleotide sequence showing $\geq 75\%$ (more preferably $\geq 85\%$, most preferably $\geq 95\%$) homology to that shown as SEQ ID NO: 3.

Preferably, the isolated DNA molecule of the seventh aspect encodes a protein comprising an amino acid sequence substantially corresponding to that shown as SEQ ID NO: 4.

20 The isolated DNA molecule of the sixth and seventh aspect may be incorporated into plasmids or expression vectors (including viral vectors), which may then be introduced into suitable bacterial, yeast, insect and mammalian host cells. Such host cells may be used to express the receptor encoded by the isolated DNA molecule.

Accordingly, in an eighth aspect, the present invention provides a 25 mammalian, insect, yeast or bacterial host cell transformed with the DNA molecule of the sixth or seventh aspect.

In a ninth aspect, the present invention provides a method of 30 producing an intracellular Na^+ receptor, comprising culturing the host cell of the eighth aspect under conditions enabling the expression of the DNA molecule and optionally recovering the expressed receptors.

35 Preferably, the host cell is mammalian, amphibian or of insect origin. Where the cell is mammalian, it is presently preferred that it be a Chinese hamster ovary (CHO) cell or human embryonic kidney 293 cell. Where the cell is of amphibian origin, it is presently preferred that it be a *Xenopus* oocyte. Finally, where the cell is of insect origin, it is presently preferred that it be an insect Sf9 cell.

In a tenth aspect, the present invention provides an intracellular Na^+ receptor designated GILT, said receptor comprising an amino acid sequence substantially corresponding to that shown as SEQ ID NO: 2, in a substantially pure form.

5 In an eleventh aspect, the present invention provides a candidate intracellular Na^+ receptor designated SCunique, said receptor comprising an amino acid sequence substantially corresponding to that shown as SEQ ID NO: 4, in a substantially pure form.

10 In a twelfth aspect, the present invention provides an antibody which specifically binds to a receptor according to the tenth or eleventh aspect. Such antibodies may be polyclonal or monoclonal and may be produced in accordance with any of the known techniques in the art.

15 The present applicants have also identified two variants of the nucleotide sequence encoding GILT (SEQ ID NO: 5 and SEQ ID NO: 6) and isolated and sequences some of the 5' non-coding sequence of the nucleotide sequence encoding SCunique (SEQ ID NO: 7). It is to be understood that the present invention extends to these additional nucleotide sequences.

20 In a thirteenth aspect, the present invention provides a method for detecting agonist or antagonist agents of the receptor of the tenth or eleventh aspect, comprising contacting said receptor, or a host cell transformed with and expressing the DNA molecule of the sixth or seventh aspect, with a test agent under conditions enabling the activation of said receptor, and detecting an increase or decrease in activity of the receptor.

25 In a further aspect, the present invention provides a nucleic acid probe/primer comprising a nucleotide sequence of 10 or more nucleotides capable of specifically hybridising to a unique sequence within a DNA molecule having a nucleotide sequence as shown as SEQ ID NO: 1 or SEQ ID NO: 3 under high stringency conditions.

30 As used herein, the term "high stringency conditions" refers to conditions that (i) employ low ionic strength and high temperature for washing, for example, 15 mM NaCl/1.5 mM sodium citrate/0.1% NaDODSO₄ at 50°C; (ii) employ during hybridisation a denaturing agent such as formamide, for example, 50% (vol/vol) formamide with 0.1% bovine serum albumin, 0.1% Ficoll, 0.1% polyvinylpyrrolidone, 50 mM sodium phosphate buffer at pH 6.5 35 with 750 mM NaCl, 75 mM sodium citrate at 42°C; or (iii) employ 50% formamide, 5 x SSC (750 mM NaCl, 75 mM sodium citrate), 50 mM sodium

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phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 µg/ml), 0.1% SDS and 10% dextran sulfate at 42°C in 0.2 x SSC (30 mM NaCl, 3 mM sodium citrate) and 0.1% SDS.

The term "substantially corresponding" as used herein in relation to 5 nucleotide sequences is intended to encompass minor variations in the nucleotide sequence which due to degeneracy in the DNA code do not result in a change in the encoded protein. Further, this term is intended to encompass other minor variations in the sequence which may be required to enhance expression in a particular system but in which the variations do not 10 result in a decrease in biological activity of the encoded protein.

The term "substantially corresponding" as used herein in relation to amino acid sequences is intended to encompass minor variations in the

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amino acid sequences which do not result in a decrease in biological activity of the encoded protein. These variations may include conservative amino acid substitutions. The substitutions envisaged are:-

5 G, A, V, I, L, M; D, E; N, Q; S, T; K, R, H; F, Y, W, H; and P, Na^α -alkylamino acids.

References to percent homology values herein are calculated by the BLAST program blastn as described by Altschul, S.F. et al., "Gapped BLAST and PSI-BLAST: a new generation of protein database search programs", Nucleic Acids Research Vol. 25, No. 17, pp 2289-3402 (1997).

10 The terms "comprise", "comprises" and "comprising" as used throughout the specification are intended to refer to the inclusion of a stated step, component or feature or group of steps, components or features with or without the inclusion of a further step, component or feature or group of steps, components or features.

15 The invention will hereinafter be further described by way of the following non-limiting example and accompanying figures.

Brief description of the accompanying figures:

20 Figure 1: Shows features of the Na^+ -dependent pH_i recovery measured with a zero Na^+ pipette solution. (A) Representative experiment with 10 mM ATP in the pipette. The bar indicates the period of readmission of 155 mM Na^+ solution to the bath. (B) Concentration-response relation for the effect of extracellular ethylisopropylamiloride (EIPA) on the Na^+ -dependent pH_i recovery. (C) The effect of modifying intracellular ATP levels.

25 Figure 2: Shows inhibition of Na^+ -dependent pH_i recovery by cytosolic Na^+ . (A) Dependency of the Na^+ -dependent pH_i recovery on pipette Na^+ . (B) The effects of inclusion of 20 mM NMDG $^+$ in the zero Na^+ pipette solution, or by buffering intracellular and extracellular Ca^{2+} to zero by the inclusion of 20 mM 1,2-bis(2-aminophenoxy)ethane-N,N,N',N'-tetraacetate (BAPTA) in the pipette solution and 1 mM EGTA in the bath solution. No Ca^{2+} was added to either solution.

30 Figure 3: Shows that the Na^+ feedback inhibition is mediated by a G protein. (A) The effect of the addition of 100 μM GDP- β -S to the pipette solution. (B) The effect of the addition of 500 ng/ml activated pertussis toxin to the pipette solution. (C) The effect of the addition to the pipette solution of antibodies directed against various G protein α -subunits [AbG_{α1,α2} = against

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C terminals of $G\alpha_i$ and $G\alpha_{i2}$; $AbG_{o,ii}$ = against C terminals of $G\alpha_o$ and $G\alpha_{op}$; AbG_{oi} = against C terminal of $G\alpha_o$; AbG_o = against N terminal of $G\alpha_o$; all 1 in 200 (vol/vol)].

Figure 4: Shows the inhibition of Na^+ feedback by intracellular amiloride. (A) Concentration-dependency of the effect of intracellular amiloride when included in 20 mM Na^+ solution. (B) The effect of the inclusion of 0.2 μM activated recombinant α -subunit of G_o (act G_o) and amiloride (10 and 30 μM) in the zero Na^+ pipette solution. AS and inact G_o denote controls in which activation solution or inactive $G\alpha_o$, respectively, were added to the pipette solution. (C) The effect of the inclusion of anti-Nedd4 antibody (A-Nd4; 1 μg purified 1gG/ml), GST-WW fusion protein (G-W; 0.3 mg/ml), GST-wild type-ubiquitin (wt; 0.3 mg/ml) or GST-dominant negative-ubiquitin (K48R) fusion protein (dn; 0.3 mg/ml) in the 20 mM Na^+ pipette solution. In A and C the broken lines indicate the mean rate of pH_i recovery observed with zero Na^+ pipette solution.

Figure 5: Shows the mechanisms of feedback inhibition by intracellular Na^+ of epithelial Na^+ channels in salivary duct (absorptive) cells (A) and Na^+ - H^+ exchange in salivary endpiece (secretory) cells (B). In each cell model, the apical membrane is on the left and the sodium pump (Na^+ , K^+ ATPase) is shown in the basolateral membrane on the right.

Example 1: Control of Na^+ - H^+ exchange in salivary secretory cells by an intracellular Na^+ receptor.

Materials and Methods.

25 **Cell Preparation.** Male Quackenbush strain mice were killed by cervical dislocation, and the mandibular glands were removed, finely minced, and incubated for 12 min in a physiological salt solution containing 1 mg/ml collagenase (Worthington type IV). The cell suspension was then dispersed by trituration and washed with fresh Na^+ rich bath solution 30 containing 145 mM NaCl, 5.5 mM KCl, 1.2 mM $MgCl_2$, 7.5 mM Na-Hepes, 7.5 mM H-Hepes, 1 mM $CaCl_2$ and 10 mM glucose; the pH was adjusted to 7.4 with NaOH. The cells were filtered through a 75- μm nylon mesh and kept on ice until required.

35 **Patch-Clamp Techniques.** A technique based on that of Demaurex and coworkers (21) was used in which the whole-cell patch-clamp technique is used to control cytosolic composition while the pH-sensitive dye, BCECF, is

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used to measure intracellular pH (pH_i). The patch-clamp techniques used were described (22), and the cells were loaded with BCECF by including it in the pipette solution. Except for the experiments summarised in Figure 1C, in which MgSO_4 replaced MgATP , pipettes were filled with solutions 5 containing 145 mM K-glutamate and Na-glutamate combined, 5 mM KCl, 5 mM Mes, 10 mM Mg-ATP, 1 mM EGTA, 40 mM sucrose, and 0.2 mM BCECF; the pH was adjusted to 6.0.

Measurement of pH_i . The equipment used to measure pH_i was as described (23). The chamber (0.3 ml) was continuously perfused with a Na^+ -free bath solution containing 145 mM N-methyl-D-glucamine (NMDG)-Cl, 5.5 mM KCl, 15 mM H-Hepes, 1.2 mM MgCl_2 , 1 mM CaCl_2 , and 10 mM glucose with a pH of 7.4. Single cells in the whole-cell configuration were voltage-clamped at -30 mV. After 3 min they were illuminated alternately at 490 and 430 nm. $\text{Na}^+ \text{-H}^+$ exchange activity was measured by reintroducing Na^+ to the bath between 2 and 3 min after the start of illumination. pH_i recovery rate was determined by fitting a linear regression to the linear phase of the pH_i recovery (i.e., between 20% and 80% of maximal recovery). Calibration of the BCECF signal was by the nigericin high- K^+ method (23).

Chemicals. Sources of chemicals and the methods for activating pertussis toxin and G protein α -subunits were as reported (24, 25). Antibodies directed against the C terminals of the α -subunits of G_i/G_{i2} , G_{i1} and G_{i3}/G_o were obtained from Calbiochem, and antibodies against the N-terminal of the α -subunit of G_o were obtained from DuPont-NEN. They were used in the pipette solution at a 1 in 200 (vol/vol) dilution of the solution 25 provided by the manufacturer. Glutathione-S-transferase (GST)-WW (G-W), GST-dominant negative-ubiquitin (K48R), and GST-wild type-ubiquitin fusion proteins were produced as described (24). The anti-Nedd4 antibody (A-Nd4) was purified IgG raised in rabbits against the C-terminal half of the protein (24, 26).

30 Results are presented as means \pm SEM. At least five cells were tested in each experimental group. Statistical significance was assessed by using Student's unpaired t test. All experiments were performed at 22°C.

Results.

Activity of $\text{Na}^+ \text{-H}^+$ exchangers was measured by a technique described 35 by Demaurex and coworkers (21) in which the whole-cell configuration of the patch-clamp technique is used to control cytosolic composition while the pH-

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sensitive dye, BCECF, measures pH_i. The cells were bathed initially in a zero Na⁺ solution so that they would be unable to oppose the acid load imposed by the pipette solution using Na⁺-dependent H⁺ transporters such as the Na⁺-H⁺ exchanger. The bath solution then was changed to one containing 155
5 mM Na⁺ so as to activate the Na⁺-H⁺ exchanger and cause pH_i to recover toward normal levels (Fig. 1A). The rate of this Na⁺-dependent pH_i recovery was used to estimate Na⁺-H⁺ exchange activity. The technique was validated by demonstrating that Na⁺-dependent pH_i recovery has features consistent
10 with its being the result of the NHE1 isoform of Na⁺-H⁺ exchanger, which predominates in salivary secretory cells. It was found that the Na⁺-dependent pH_i recovery was highly sensitive to the amiloride analog, ethylisopropylamiloride (Fig. 1B), and that the recovery depended on the presence of ATP (21), being inactivated when intracellular ATP was depleted
15 by treatment with 2-deoxy-D-glucose (5 mM) and oligomycin (5 µg/ml; Fig. 1C).

It was demonstrated that the rate of the Na⁺-dependent pH_i recovery declined with increasing pipette Na⁺ concentration (Fig. 2A) in a manner similar to that described in sheep F2 Purkinje fibres (27). This inhibition evidently was caused by increased [Na⁺]_i because it could not be reproduced
20 by the large organic cation, NMDG⁺ (Fig. 2B). Because intracellular free Ca²⁺ is known to regulate Na⁺-H⁺ exchangers (28), an investigation was made to determine whether a change in free intracellular Ca²⁺ concentration could mediate this phenomenon. It was found that buffering cytosolic and
25 extracellular Ca²⁺ to nominal zero did not alter the effect of increased [Na⁺]_i (Fig. 2B).

An investigation was also made to determine the mechanism by which [Na⁺]_i controls the activity of the Na⁺-H⁺ exchanger. It was found that inclusion of the pipette solution of 100 µM GDP-β-S (which competitively inhibits the binding of GTP by G proteins; ref. (29) or of 500 ng/ml activated
30 pertussis toxin (which ADP ribosylates G proteins of the G_i and G_o classes so as to prevent their interaction with receptors; ref. (30), reversed the inhibitory effect of 20 mM Na⁺ (Fig. 3A and B). The ability of these agents to overcome the inhibitory effect of raised intracellular Na⁺ completely without altering the electrochemical gradient for Na⁺ indicates that the inhibition is
35 not caused by a decreased electrochemical driving force for Na⁺-H⁺ exchange. Rather, it must be caused by a G protein-mediated feedback

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pathway. In this regard, it was further found that inclusion in the pipette solution of antibodies directed against the α -subunit of the G_o protein, which is known to be expressed in salivary endpiece cells (31), abolished the inhibitory effect of 20 mM Na^+ . In contrast, antibodies directed against the α -subunits of G_{i1} , G_{i2} , and G_{s1} were without effect (Fig. 3C).

In the absorptive cell of the salivary duct, $[Na^+]_i$ is sensed by a receptor the effect of which is mediated by G_o (10). This receptor is blocked by amiloride and amiloride analogs such as dimethylamiloride and benzimidazolylguanidinium, thus explaining the ability of these agents to stimulate Na^+ channel activity. It was found that the inclusion of amiloride in the pipette solution reversed the inhibitory effect of 20 mM Na^+ (Fig. 4A). Further, it was found that the inclusion of the activated α -subunit of G_o in the zero Na^+ pipette solution (Fig. 4B) inhibited the Na^+ - H^+ exchanger and that the inclusion of as much as 30 μ M amiloride in the pipette solution was unable to overcome this inhibition (Fig. 4B). Thus, amiloride exerts its inhibitory action upstream of G_o , presumably at the putative receptor for intracellular Na^+ .

Discussion.

It has been previously shown that $[Na^+]_i$ and the G protein, G_o , regulate the activity of the epithelial Na^+ channel in the duct cells of the mouse mandibular gland via Nedd4 (24), a ubiquitin-protein ligase that is believed to bind to Na^+ channels and regulate their activity by ubiquitinating them (12, 13). Here, it was found that feedback inhibition of the Na^+ - H^+ exchanger was not prevented by inclusion in the pipette solution of an antibody directed against Nedd4 or of a fusion protein composed of GST and the three WW-domains of mouse Nedd4 (GST-W), which acts as a dominant negative mutant of Nedd4 (Fig. 4C). This finding is consistent with the low level of expression of Nedd4 in endpiece cells (24). Feedback inhibition was blocked, however, by inclusion of a dominant negative mutant of ubiquitin (K48R) (24) in the pipette solution (Fig. 4C), indicating that feedback regulation of the exchanger nevertheless is mediated by ubiquitination. Because our preliminary data show that NHE1 transfected into COS cells is ubiquitinated (data not shown), the findings indicate that feedback regulation of NHE1 is mediated by ubiquitination of the exchanger protein. The control system then would resemble the control of surface expression of epithelial Na^+ channels by ubiquitination of the channel protein catalysed by Nedd4.

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It cannot be excluded however, that the inactivation of NHE1 produced by Na⁺ feedback is the result of ubiquitination of a protein associated with the exchanger, as recently has been proposed for the control of the growth hormone receptor by ubiquitination (32). Whatever the mechanism, the 5 present findings taken together with the finding that activity of epithelial Na⁺ channels can be rapidly down-regulated by ubiquitination suggest that ubiquitination may be a general mechanism for the rapid control of membrane transport protein activity.

10 **Example 2: Prevention of the progression of diabetic nephropathy and other forms of chronic renal failure by 6-iodoamiloride.**

Materials and methods.

20 mg 6-iodoamiloride tablets may be formulated and taken orally at a dosage of one or two every 6 hours.

15 **Discussion.**

6-iodoamiloride acts by blocking the intracellular Na⁺ receptor that controls NHE1 and other sodium-dependent transporters as well as mediating the normal cellular responses to increased intracellular sodium concentration (which include release of cytokines and increased cell growth and proliferation (44). In this way, cytokine release and cellular proliferation caused by increased intracellular sodium can be treated with 6-iodoamiloride to prevent the cytokine release and cell growth and proliferation that lead to progression of renal failure.

25 **Example 3: Treatment of cells with reduced Na⁺ transport with recombinant adenovirus.**

Materials and methods.

30 **Recombinant adenovirus.** Recombinant adenovirus including an expressible gene encoding the Na⁺ receptor, GILT may be prepared by routine molecular biology techniques (33). Particularly, the clone encoding GILT (SEQ ID NO:1) may be ligated to a suitable mammalian promoter sequence (e.g. CMV (45)) and inserted into a suitable vector for the transfer, by homologous recombination, of the recombinant GILT gene into an adenovirus as described by He et al. (46).

35 **Administration.** The recombinant adenovirus may be formulated and administered in accordance with known methods in the art. In particular,

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the recombinant adenovirus may be formulated for administration as a nasal spray or intrabronchial spray or given intravenously (47, 48) or direct injections of muscle or of organs (49). With administration to the respiratory tract (50), the recombinant adenovirus will preferably be administered at a 5 dose of 10^9 plaque forming units (pfu) at intervals between 2 and 4 weeks.

Discussion.

Upon infection of host diseased cells, the adenovirus will bring about the expression of functional GILT protein to decrease Na^+ transport and restore cytosolic ion composition to substantially that of corresponding 10 healthy cells.

Example 4: Prevention of the progression of chronic hypoxic pulmonary hypertension and other forms of pulmonary hypertension by 6-iodoamiloride and other inhibitors of the sodium receptor.

Materials and methods.

20mg 6-iodoamiloride tablets may be formulated and taken orally at a dosage of 1 or 2 every 6 hours.

Discussion.

6-iodoamiloride acts by blocking the intracellular Na^+ receptor that 20 controls NHE1 and other sodium-dependent transporters as well as mediating the normal cellular responses to increased intracellular sodium concentration (which include release of cytokines and increased cell growth and proliferation (44). In this way, cytokine release and cellular proliferation 25 caused by increased intracellular sodium can be treated with 6-iodoamiloride to prevent the cytokine release and cell growth and proliferation that lead to progression of pulmonary hypertension due to chronic hypoxia (51).

It will be appreciated by persons skilled in the art that numerous 30 variations and/or modifications may be made to the invention as shown in the specific embodiments without departing from the spirit or scope of the invention as broadly described. The present embodiments are, therefore, to be considered in all respects as illustrative and not restrictive.

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References:

1. Cui Y, Su YR, Rutkowski M, Reif M, Menon AG, Pun RY (1997) Loss of protein kinase C inhibitor in the beta T594M variant of the amiloride-sensitive Na^+ channel. *Proc Natl Acad Sci USA* 94:9962-9966.
5. Lifton RP (1996) Molecular genetics of human blood pressure variation. *Science* 272:676-680.
10. 3. Sweeney FP, Siczkowski M, Davies JE, Quinn PA, McDonald J, Krowlewska AS, Ng LL (1995) Phosphorylation and activity of Na^+/H^+ exchanger isoform 1 of immortalized lymphoblasts in diabetic nephropathy. *Diabetes* 44:1180-1185.
15. 4. Gaspardone A, Ferri C, Crea F, F Versaci, Tomai F, Santucci A, Chiarieillo L, Gioffre PA (1998) Enhanced activity of sodium-lithium countertransport in patients with cardiac syndrome X: a potential link between cardiac and metabolic syndrome X. *J. Am Coll Cardiol.* 32:2031-2034.
20. 5. Neves PI, Faisca M, Gomes V, Cacodcar S, Bernardo I, Anunciada AI, Viegas E, Martins H, da Silva AM (1996) Risk factors for left ventricular hypertrophy: role of Na^+/Li^+ countertransport. *Kidney Int.* 55:S160-S162.
25. 6. Quinn D.A., Du H.K., Thompson B.T., Hales C.A. (1998) Amiloride analogs inhibit chronic hypoxic pulmonary hypertension. *Amer. J. Resp. Critical Care Med.* 157:1263-1268.
30. 7. Stutts MJ, Canessa CM, Olsen JC, Hanrick M, Cohn JA, Rossier BC, Boucher RC (1995) CFTR as a cAMP-dependent regulator of sodium channels. *Science* 269: 847-850.
35. 8. Dinudom A, Harvey KF, Komwatana P, Young JA, Kumar S, Cook DI (1998) Nedd4 mediates control of an epithelial Na^+ channel in salivary duct cells by cytosolic Na^+ . *Proc Natl Acad Sci USA* 95:7169-7173.

WO 01/12805

PCT/AU00/00980

17

9. Kellenberger S, Gautschi I, Rossier BC, Schild L (1998) Mutations causing Liddle syndrome reduce sodium-dependent downregulation of the epithelial sodium channel in the *Xenopus oocyte* expression system. *J Clin Invest* 101:2741-2750.
10. Komwatana P, Dinudom A, Young J. A. and Cook D. I. (1998) *J. Membr. Biol* 162:225-232.
- 10 11. Komwatana P, Dinudom A, Young J. A. and Cook, D. I (1996) *Proc Natl Acad Sci USA* 93:8107-8111.
12. Staub O, Dho S, Henry P. C., Correa J, Ishikawa T, McGlade J, and Rotin, D. (1996) *EMBO J.* 15, 2371-2380.
- 15 13. Staub O, Gautschi I, Ishikawa T, Breitschopf K, Ciecanover, A, Schild L, and Rotin D. (1997) *EMBO J.* 16:6325-6336.
14. Garty H., and Palmer L.G. (1997) *Physiol Rev* 77:359-396.
- 20 15. Cussac D., Vidal M., Leprince C., Lin W.Q., Cornille F., Tiraboschi G., Roque B.P. and Garbay C. (1999) A Sos-derived peptidimer blocks the Ras signalling pathway by binding both the Grb2 SH3 domains and displays antiproliferative activity. *FASEB Journal* 13:31-38.
- 25 16. Hollinger E.P., Chittenden T. and Lutz R.J. (1999) Bak BH3 peptide antagonises Bcl-x2 function and induces apoptosis through cytochrome C-independent activation of caspases. *J. Biol. Chem.* 274:13298-13304.
- 30 17. Lang J., Nishimoto I., Okamoto T., Regazzi R., Kiraly C., Weller U., Wollheim C.B. (1995) Direct control of exocytosis by receptor-mediated activation of the heterotrimeric GTPases G_i and G_o or by expression of their active G α subunits. *EMBO Journal* 14:3635-3644.

35

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PCT/AU00/00980

18

18. Lin P., Le-Niculescu H., Hosfmeister R., McCaffery J.M., Jin M., Hennemann H., McQuistan T., De Vries L., Farquhar M.G. (1998) The mammalian calcium-binding protein, nucleobindin (CALNUC), is a Golgi resident protein. *J. Cell. Biol.* 141:1515-1527.
- 5 19. Berman D.M., Wilkie T.M., Gilman A.G. (1998) GAIP and RSG4 are GTPase-activating proteins for the G_i subfamily of G protein alpha subunits. *Cell* 86:445-452.
- 10 20. Jordan J.D., Carey K.D., Stork P.J.S., Iyengar R. (1999) Modulation of Rap activity by direct interaction of Galphao with Rap1GTPase-activating protein. *J. Biol. Chem.* 274:21507-21510.
- 15 21. Demaurex N, Romanek RR., Orlowski J, and Grinstein S. (1997) *J. Gen. Physiol* 109:117-128.
22. Dinudom A, Young JA., and Cook DI. (1993) *J. Membr. Biol.* 135, 289-295.
- 20 23. Steward MC., Poronnik P, and Cook DI. (1996) *J. Physiol. (London)* 494:819-830.
24. Dinudom A, Harvey K. F., Komwatana, P, Young J. A., Kumar, S, and Cook D. I. (1998) *Proc Natl Acad Sci USA* 95:7169-7173.
- 25 25. Dinudom A, Komwatana P, Young JA. and Cook DL. (1995) *J. Physiol. (London)* 487:549-555.
26. Kumar S, Harvey KF, Kinoshita M, Copland NG, Noda M, and Jenkins NA (1997) *Genomics* 40:435-443.
- 30 27. Wu ML and Vaughan Jones RD (1997) *J. Mol Cell Cardiol* 29:1131-1140.
28. Orlowski J and Grinstein S. (1997) *J. Biol Chem* 272:22373-22376.

35

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PCT/AU00/00986

19

29. Eckstein F., Cassel D., Levkovitz H., Lowe M. and Sclinger Z. (1979) J. Biol Chem 254:9829-9834.
30. Katada T. and Ui M. (1982) J. Biol Chem 257:7210-7216.
- 5 31. Watson EL, Olver C., D'Silva N., and Belton C. M. (1994) J. Histochem Cytochem 42:41-47.
- 10 32. Govers R, ten Broeke, T, van Kerkhof P, Schwartz AL, and Strous GJ. (1999) EMBO J. 18:28-36.
- 15 33. Sambrook J., Fritsch E.S. and Maniatis T. (1989) Molecular Cloning: a laboratory manual, Second Edition, Cold Spring Harbor Laboratory Press, USA.
- 20 34. Ishibashi H, Dinudom A, Harvey KF, Kumar S, Young JA and Cook DI (1999) Proc. Natl. Acad. Sci. USA 96:9949-9953.
35. Counillon L. and Pouysegur J. (2000) J. Biol. Chem. 275:1-4.
- 25 36. Orlowski J. and Grinstein S. (1997) J. Biol. Chem. 272:22373-22376.
37. Romero M.F. and Boron W. (1999) Annu. Rev. Physiol. 61:699-723.
- 30 38. Haas M. and Forbush B 3rd (1998) J. Bioenerg. Biomembr. 30:161-172.
39. Komwatana P., Dinudom A., Young J.A. and Cook D.I. (1996) Proc. Natl. Acad. Sci. USA 93:8107-8111.
40. 40. Friessmuth M., Beindl W., Nickel P., Ijzerman A.P., Hohenegger M. and Nanoff C. (1996) Mol. Pharmacol. 49:602-611.
41. Gautschi I., Ishikawa T., Breitschopf K., Ciechanover A., Schild L. and Rotin D. (1997) EMBO J 16:6325-6336.

WO 01/12805

PCT/AU00/00980

20

42. Volchuk A., Narine S., Foster L.J., Grabs D., De Camilli P. and Klip A. (1998) *Biol. Chem.* 273:8169-8176.
43. Dinudom A., Komwatana P., Young J.A. and Cook D.L (1995) *J. Physiol. (london)* 487:549-555.
44. Stanton B.A. and Kaissling B. (1989) *Am. J. Physiol.* 257:F1-F10.
45. Cummings MM, O'Mulane LM, Barden JA, Cook DI & Poronnik P. (2000) *Cell Calcium* 27: 247-255.
46. He TC, Zhou S, da Costa LT, Yu J, Kinzler KW & Vogelstein BA. A simplified system for generating recombinant adenoviruses. *Proc. Natl Acad. Sci. USA* 95: 2509-2514, 1998.
47. Nemunatis J, Swisher SG, Timmons T, Connors D, Mack M, Doerkson L, Weill D, Wait J, Lawrence DD, Kemp BL, Fossella F, Glisson BS, Hong WK, Khuri FR, Kurie JM, Lee JJ, Lee JS, Nguyen DM, Neshitt JC, Perez-Soler R, Pisters KM, Putnam JB, Richli WR, Shin DM, Walsh GL et al. Adenovirus-mediated p53 gene transfer in sequence with cisplatin to tumours of patients with non-small-cell lung cancer. *J. Clin. Oncol.* 18: 609-622, 2000.
48. Dobrzynski E, Yoshida H, Chao J & Chao L. Adenovirus-mediated kallikrein gene delivery attenuates hypertension and protects against renal injury in deoxycorticosterone-salt rat. *Immunopharmacology* 44: 57-65, 1999.
49. Rosengart TK, Lee LY, Patel SR, Kligfield PD, Okin PM, Hackett NR, Isom OW & Crystal RG. Six-month assessment of a phase I trial of angiogenic gene therapy for the treatment of coronary artery disease using direct intramyocardial administration of an adenovirus vector expressing VEGF121 cDNA. *Ann. Surg.* 230: 466-470, 1999.
50. Harvey BG, Leopold PL, Hackett NR, Grasso TM, Williams PM, Tucker AL, Kaner RJ, Ferris B, Gonda I, Sweeney TD, Ramalingam R, Kovesdi

WO 01/12805

PCT/AU00/00980

21

I, Shak S & Crystal RG. Airway epithelial CFTR mRNA expression in cystic fibrosis patients after repetitive administration of a recombinant adenovirus. *J. Clin. Invest.* 104: 1245-1255, 1999.

5 51. Quinn DA, Du H-K, Thompson BT & Hales CA. Amiloride analogues inhibit chronic hypoxic pulmonary hypertension. *Am. J. Resp. Crit. Care Med.* 157: 1263-1268, 1998.

Claims:

1. A method of treatment of a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from 5 inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has reduced activity, the method comprising administering to a subject having said disease an effective amount of an agent 10 that blocks said inhibitory feedback mechanism and thereby substantially restores the ion composition of the cytosol in said diseased cells to that which is found in corresponding cells from healthy tissue.
2. A method according to claim 1, wherein the Na^+ transport protein is selected from the group consisting of Na^+ - H^+ exchanger 1 (NHE1), Na^+ - H^+ 15 exchanger 2 (NHE2), Na^+ - H^+ exchanger 3 (NHE3), the Na^+ - HCO_3^- cotransporter and the Na^+ - K^+ - 2Cl^- cotransporter.
3. A method according to claim 1 or 2, wherein the agent is selected from 20 amiloride and amiloride analogs.
4. A method according to claim 1 or 2, wherein said agent is a G-protein inhibitor.
5. A method according to claim 4, wherein the G-protein inhibitor is 25 selected from GDP- β -S and NFD23.
6. A method according to claim 1 or 2, wherein said agent is a ubiquitin protein ligase inhibitor.
7. A method according to claim 6, wherein the ubiquitin protein ligase inhibitor is selected from dominant negative mutants of ubiquitin and agents 30 that prevent binding of ubiquitin protein ligase to Na^+ transport proteins.
8. A method of treatment of a human disease which is characterised by 35 abnormal cytosolic ion composition in diseased cells resulting from the loss

of or inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has over activity, the method comprising administering to a subject having said disease an effective amount of an agent that substantially restores the ion composition of the cytosol in said diseased cells to that which is found in corresponding cells from healthy tissue.

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9. A method according to claim 8, wherein said agent is selected from gene therapy agents, intracellular Na^+ receptor activators, G-protein activators, ubiquitin ligase activators and endocytosis triggering agents.

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10. A method according to claim 9, wherein said gene therapy agent is an adenovirus including a nucleotide sequence encoding a non-mutated protein which participates in the Na^+ transport protein inhibitory feedback mechanism.

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11. A method according to claim 9, wherein said intracellular Na^+ receptor activator is selected from guanidium and guanidium analogs.

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12. A method according to claim 9, wherein said G-protein activator is selected from GDP- γ -S and receptor mimetic peptides.

13. A method of diagnosis of a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from the loss 25 of or inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has reduced or over activity, the method comprising isolating from a subject suspected of having said disease a sample of cells and assessing said sample of cells for reduced or over activity of said 30 Na^+ transport protein resulting from the loss of or inappropriate activity of said inhibitory feedback mechanism or otherwise assessing said sample of cells for the loss of or inappropriate activity of said inhibitory feedback mechanism.

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14. A method according to claim 13, wherein said Na^+ transport protein is selected from the group consisting of Na^+ - H^+ exchanger 1 (NHE1), Na^+ - H^+ exchanger 2 (NHE2), Na^+ - H^+ exchanger 3 (NHE3), the Na^+ - HCO_3^- cotransporter and the Na^+ K^+ 2Cl cotransporter.

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15. A method according to claim 13 or 14, wherein the sample of cells is a sample of lymphocytes.

16. A method according to any one of claims 13 to 15, wherein the sample of cells is assessed for reduced or over activity of the Na^+ transport protein by determining the rate of Na^+ -dependent intracellular pH (pH_i) recovery and comparing said rate against similarly measured rates from cells from healthy tissue isolated from said subject or a control subject(s).

15 17. A method of diagnosis of a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from the loss of or inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has reduced or over activity, the method comprising isolating from a subject suspected of having said disease a sample of cells and assessing said sample of cells for over or under expression of the Na^+ transport protein or another protein participating in the Na^+ transport protein inhibitory feedback mechanism.

25 18. A method of diagnosis of a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from the loss of or inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has reduced or over activity, the method comprising isolating a genomic DNA sample from a subject suspected of having said disease and assessing said sample for the presence of a gene encoding a mutated product causative of said reduced or over activity of said Na^+ transport protein.

19. A method of assessing a subject for a predisposition to a human disease which is characterised by abnormal cytosolic ion composition in diseased cells resulting from the loss of or inappropriate activity of an inhibitory feedback mechanism controlling the activity of an Na^+ transport protein other than an epithelial Na^+ channel such that the Na^+ transport protein has reduced or over activity, the method comprising isolating a genomic DNA sample from a subject and assessing said sample for the presence of a gene encoding a mutated product causative of said reduced or over activity of said Na^+ transport protein.

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20. A method according to any one of claims 17-19, wherein said Na^+ transport protein is selected from the group consisting of Na^+ - H^+ exchanger 1 (NHE1), Na^+ - H^+ exchanger 2 (NHE2), Na^+ - H^+ exchanger 3 (NHE3), the Na^+ - HCO_3^- cotransporter and the Na^+ - K^+ - 2Cl^- cotransporter.

21. A method according to any one of claims 1 to 20, wherein the said human disease is selected from hypertension, renal failure, cardiac hypertrophy and cardiological syndrome X.

22. An isolated DNA molecule encoding an intracellular Na^+ receptor designated GILT, said DNA molecule comprising a nucleotide sequence showing $\geq 75\%$ homology to the nucleotide sequence shown as SEQ ID NO: 1.

23. A DNA molecule according to claim 22, wherein said DNA molecule comprises a nucleotide sequence showing $\geq 85\%$ homology to the nucleotide sequence shown as SEQ ID NO: 1.

24. A DNA molecule according to claim 22, wherein said DNA molecule comprises a nucleotide sequence showing $\geq 95\%$ homology to the nucleotide sequence shown as SEQ ID NO: 1.

25. A DNA molecule according to claim 22, wherein said DNA molecule comprises a nucleotide sequence substantially corresponding to that shown as SEQ ID NO: 1.

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26. A DNA molecule according to claim 22, wherein said DNA molecule encodes a protein comprising an amino acid sequence substantially corresponding to that shown as SEQ ID NO: 2.

5 27. An isolated DNA molecule encoding an intracellular Na^+ receptor designated SCunique, said DNA molecule comprising a nucleotide sequence showing $\geq 75\%$ homology to the nucleotide sequence shown as SEQ ID NO: 3.

10 28. A DNA molecule according to claim 27, wherein said DNA molecule comprises a nucleotide sequence showing $\geq 85\%$ homology to the nucleotide sequence shown as SEQ ID NO: 3.

15 29. A DNA molecule according to claim 27, wherein said DNA molecule comprises a nucleotide sequence showing $\geq 95\%$ homology to the nucleotide sequence shown as SEQ ID NO: 3.

30. A DNA molecule according to claim 27, wherein said DNA molecule comprises a nucleotide sequence substantially corresponding to that shown as SEQ ID NO: 3.

20 31. A DNA molecule according to claim 27, wherein said DNA molecule encodes a protein comprising an amino acid sequence substantially corresponding to that shown as SEQ ID NO: 4.

25 32. A host cell transformed with a DNA molecule according to any one of claims 22 to 31.

30 33. A method of producing an intracellular Na^+ receptor, comprising culturing the host cell of claim 32 under conditions enabling the expression of said DNA molecule and optionally recovering the expressed receptors.

35 34. An intracellular Na^+ receptor designated GILT, said receptor comprising an amino acid sequence substantially corresponding to that shown as SEQ ID NO: 2, in a substantially pure form.

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35. An intracellular Na^+ receptor designated SCunique, said receptor comprising an amino acid sequence substantially corresponding to that shown as SEQ ID NO: 4, in a substantially pure form.

5 36. An antibody which specifically binds to a receptor according to claim 34 or 35.

10 37. A method for detecting agonist or antagonist agents of the receptor of claim 34 or 35, wherein said method comprises contacting said receptor, or a host cell transformed with and expressing the DNA molecule of any one of claims 22 to 31, with a test agent under conditions enabling the activation of said receptor, and detecting an increase or decrease in activity of the receptor.

15 38. A nucleic acid probe or primer comprising a nucleotide sequence of 10 or more nucleotides, wherein said probe or primer specifically hybridises to a unique sequence within the DNA molecule of claim 25 or 30 under high stringency conditions.

20 39. An isolated DNA molecule comprising a nucleotide sequence substantially corresponding to that shown as SEQ ID NO: 5.

40. An isolated DNA molecule comprising a nucleotide sequence substantially corresponding to that shown as SEQ ID NO: 6.

25 41. An isolated DNA molecule comprising a nucleotide sequence substantially corresponding to that shown as SEQ ID NO: 7.

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(54) Title: METHODS FOR DIAGNOSIS AND TREATMENT OF HUMAN DISEASES INCLUDING HYPERTENSION

(57) Abstract: Methods for diagnosis and treatment of human disease, particularly human disease characterised by abnormal cytosolic ion composition in diseased cells resulting from reduced or over activity of Na⁺ transport proteins (e.g. hypertension), are disclosed. Additionally, the specification discloses novel Na⁺ receptors and isolated DNA molecules encoding same.

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Figure 1

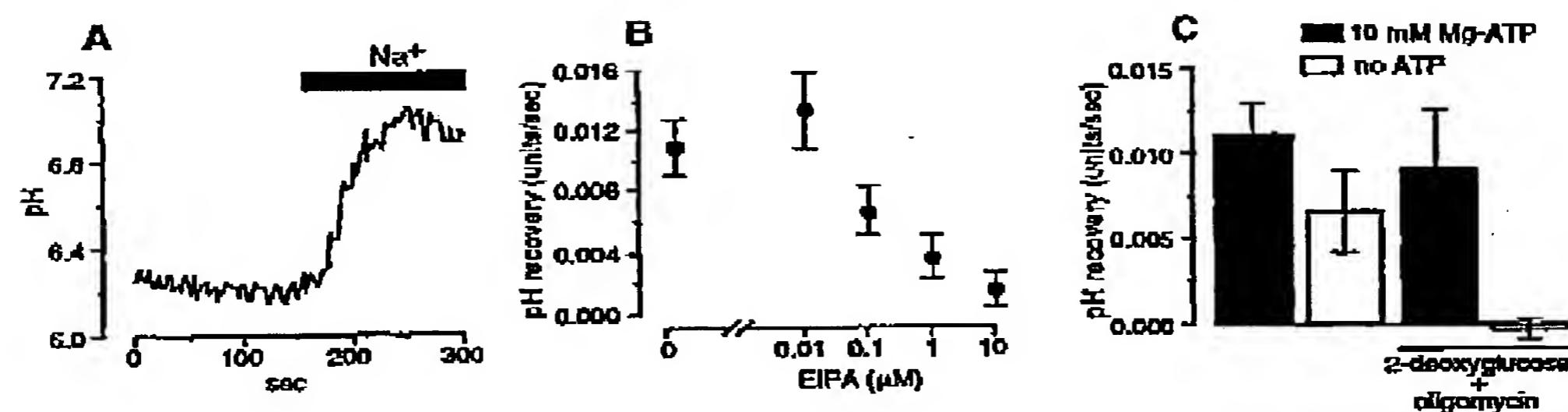
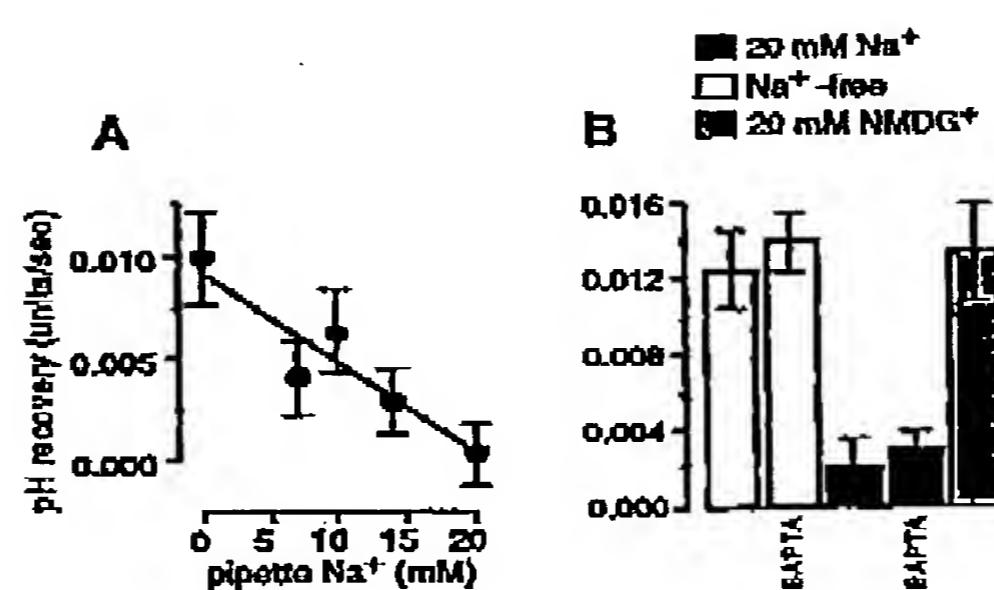
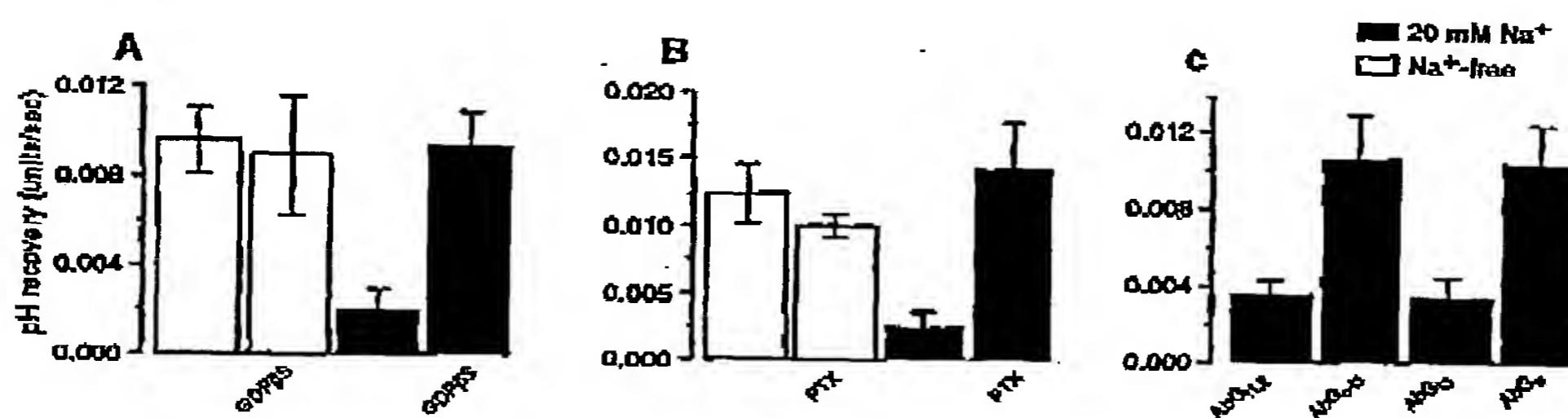


Figure 2



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Figure 3

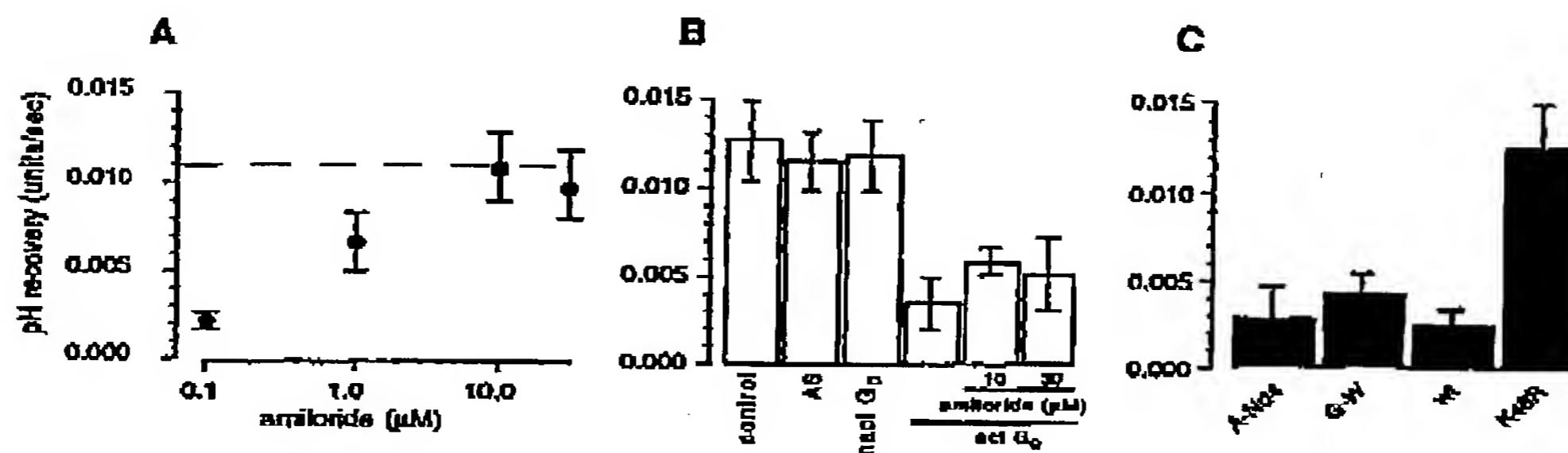


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Figure 4

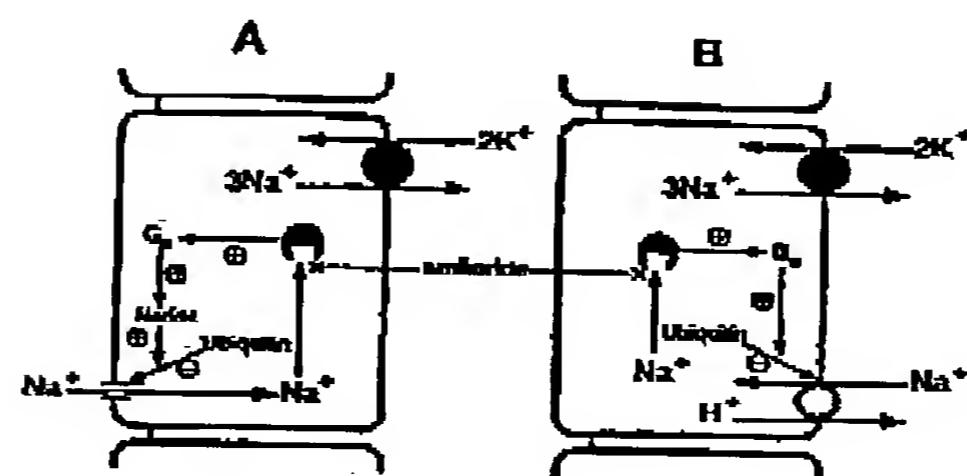


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Figure 5



DECLARATION, POWER OF ATTORNEY AND PETITION

As below named inventors, we hereby declare that:

Our residence, post office and citizenship are as stated below next to our names,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original first and joint inventor (if plural names are listed below) of the subject matter claimed and for which a patent is sought on the invention entitled: **Methods for diagnosis and treatment of human diseases including hypertension**

the specification of which

is attached hereto was filed on **16 August 2000** as Application No. **PCT/AU00/00980** and was amended on (if applicable).

We hereby state that we have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

We acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, Section 1.56(a)

We hereby claim foreign priority benefits under Title 35, United States Code, Section 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)			Priority Claimed	
[Number]	[Country]	[Day/Month/Year Filed]	Yes	No
PQ2239	Australia	16 August 1999	<input checked="" type="checkbox"/>	<input type="checkbox"/>
PCT/AU00/00980	PCT	16 August 2000	<input checked="" type="checkbox"/>	<input type="checkbox"/>
			<input type="checkbox"/>	<input type="checkbox"/>

We hereby claim the benefit under Title 35, United States Code, Section 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, Section 112, we acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, Section 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application.

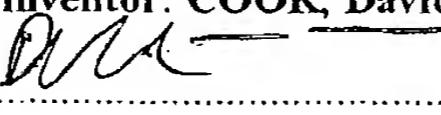
[Application Serial no]	[Filing Date]	[Status: patented, pending, abandoned]

We hereby declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that wilful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such wilful false statements may jeopardize the validity of the application or any patent issued thereon.

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with full power of substitution and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith, and all future correspondence should be addressed to them.

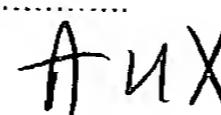
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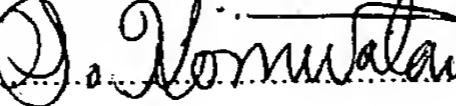
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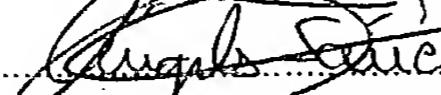
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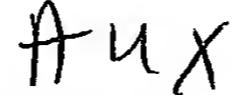
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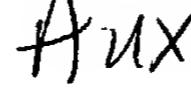
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